



Analysis of Water-Soluble Vitamins Using Quasar SPP HILIC Column

Introduction

In 1906, English biochemist Sir Frederick Gowland Hopkins also discovered that certain food factors were important to health. All the vitamins we recognise today were discovered during the early

20th century.¹ There are two groups of vitamins, fat soluble and water soluble. Both types are regarded as essential for normal growth and our overall well being.

Vitamin B₂ (Riboflavin) and vitamin B₃ (Niacinamide) are water soluble vitamins, naturally occurring in a number of food stuffs including milk and eggs. Both vitamins are used as dietary supplements and can help lower cholesterol, prevent migraines.²

Hydrophilic interaction liquid chromatography (HILIC), as first suggested by Alpert in 1990 provides an alternative approach to effectively separate small polar compounds, on polar stationary phases, which are not well retained under reversed phase conditions.³ Water is the strongest solvent in HILIC and as mobile phases typically contain low levels of water the resulting methods are ideally suited for MS applications.

This application brief illustrates the efficient separation of vitamin B₂ and B₃, Figure 1, applying the HILIC mode of separation using the Quasar SPP HILIC column.

Experimental Conditions

Method Parameters

All HPLC method parameters are shown in Table 1.

Table 1. HPLC Method Parameters.

Quasar SPP HILIC	150 mm	4.6 mm	5 µm	N9308960
Mobile Phase	H ₂ O: ACN, 5:95			
Flow Rate	1 mL/min			
Temp	30 °C			
Wavelength	254 nm			
Injection Vol.	5 µl			
Analyte	1. Vitamin B ₂ 2. Vitamin B ₃			

Solvents and Samples

All solvents were HPLC grade and samples were filtered using a 0.45 µm nylon filter, P/N 02542880.

Results and Discussion

Any polar chromatographic surface can be used for HILIC separations. Typical HILIC stationary phases consist of classical bare silica or silica modified with polar functional groups.

HILIC is a hybrid of normal phase (NP), reverse phase (RP) and ion chromatography techniques, (Figure 2). The eluents of RP combined with the stationary phases of NP and charged analytes of ion chromatography yield the basis of HILIC. The mechanism of separation has been the subject of much discussion in the literature

however it is generally agreed that a water-rich layer forms on the surface of the polar stationary phase vs. the water-deficient mobile phase, creating liquid/liquid partitioning. However, the separation mechanism is more complex than partitioning alone, with dipole-dipole and electrostatic interactions also contributing to retention.

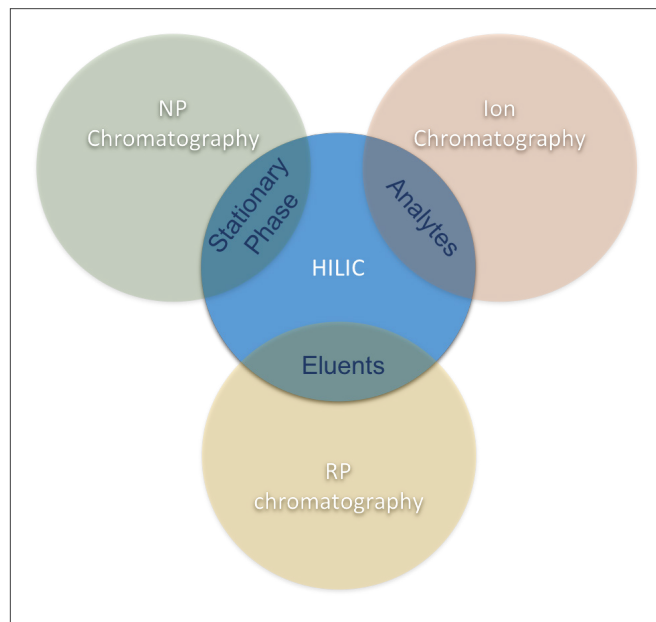


Figure 2. HILIC a hybrid of techniques.

The more polar compounds will have a stronger interaction with the stationary aqueous layer and are therefore retained longer than the less polar compounds. The elution order opposite to that observed in reverse phase HPLC.

Vitamins B₂ and B₃ contain several polar functionalities, which can also be in a charged state, and results in little retention using an alkyl chain bonded phase and traditional RP separations. So they are ideal candidates for HILIC separations. As Figure 3 illustrates, a good retention of these polar compounds is readily achieved in the HILIC mode, with vitamin B₂ eluting after vitamin B₃, as it is more polar.

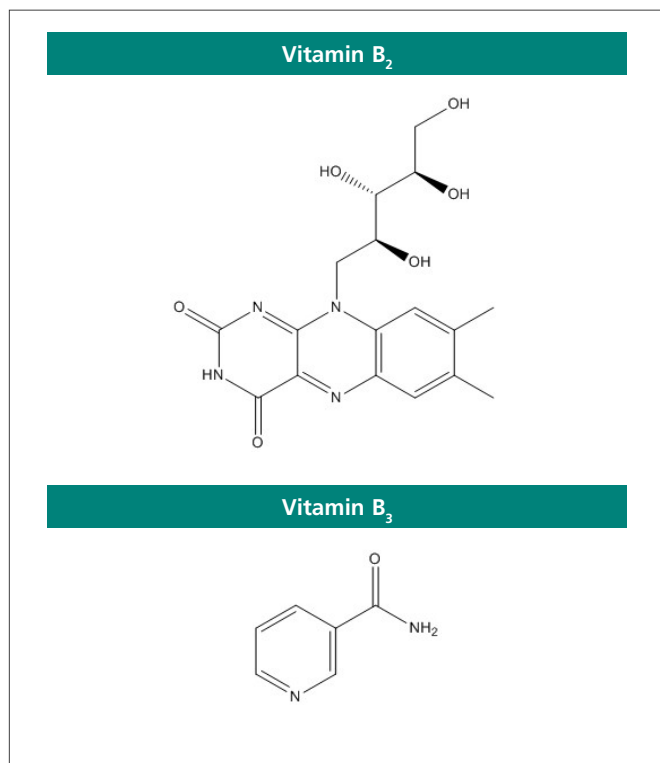


Figure 1. Chemical structures of vitamin B₂ and B₃.

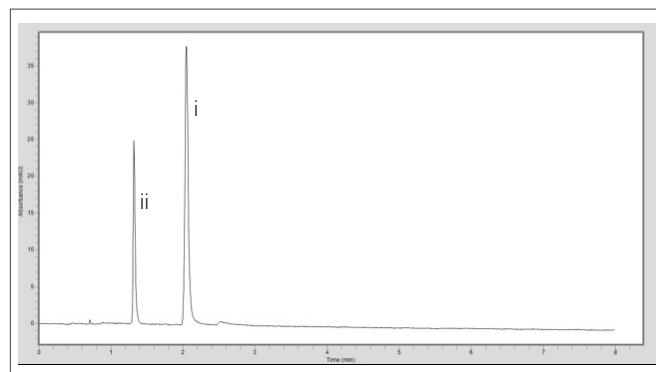


Figure 3. HPLC Analysis of (i) vitamin B₂ and (ii) vitamin B₃.

Conclusion

- Quasar SPP HILIC phase is an unbonded silica phase for use in the HILIC mode of separation.
- It offers increased retention of polar compounds over RP separations and vitamins B₂ and B₃ are well retained.
- The HILIC mode offers enhanced MS sensitivity due to the high organic content of the mobile phase.
- With a solid core and outer porous silica coating of the SPP phase you can realise up to a 50% improvement in cost and efficiency over traditional porous silica columns.

References

1. <https://www.vitamins-nutrition.org/vitamins/history-vitamins.html>
2. "Riboflavin". Drugs.com, The American Society of Health-System Pharmacists. 1 August 2018. Archived from the original on 30 December 2016. Retrieved 7 November 2018.
3. Alpert AJ. J Chromatogr A. 1990;499:177–196

Consumables

Phase	Length (mm)	I.D. (mm)	µm	Part
Quasar SPP HILIC	150	4.6	2.6	N9308919
Quasar SPP HILIC	100	4.6	2.6	N9308920
Quasar SPP HILIC	50	4.6	2.6	N9308921
Quasar SPP HILIC	150	3	2.6	N9308922
Quasar SPP HILIC	100	3	2.6	N9308923
Quasar SPP HILIC	50	3	2.6	N9308924
Quasar SPP HILIC	150	2.1	2.6	N9308925
Quasar SPP HILIC	100	2.1	2.6	N9308926
Quasar SPP HILIC	50	2.1	2.6	N9308927
Quasar SPP HILIC	150	4.6	5	N9308960
Quasar SPP HILIC	100	4.6	5	N9308961
Quasar SPP HILIC	50	4.6	5	N9308962
Quasar SSP HILIC Guard Cartridge (3/pack)	10	3	2.6	N9308994
Quasar SSP HILIC Guard Cartridge (3/pack)	10	3	5	N9308995
Quasar Guard Cartridge Holder	-	-	-	N9306876
Nylon Filters	-	-	-	02542880